Introduction

Thymic epithelial tumors (TETs) originate in the thymus and include thymomas (Ts), thymic carcinomas (TCs), and thymic neuroendocrine tumors (TNETs) (1,2). Although TETs are one of the rarest malignancies, they are the most common malignancies of the anterior mediastinum. The incidence of TETs is estimated to be 3.93 cases/million in China (3), but TETs are significantly rarer in North
America, in which the incidence is 1.5 cases/million (4). This difference in incidence is of interest, as it may indicate the genetic differences between ethnicities (4).

According to the World Health Organization (WHO), Ts are further classified into 6 different histological subtypes (A, AB, B1, B2, and B3 of Ts, and C represents TC) (5). The subtypes are evaluated by morphology and lymphocyte/epithelial cell ratio. Although TCs are type C in the WHO classification, they are significantly different from Ts. TCs are rarer but much more aggressive. The 5-year survival rate for Ts is approximately 90% (6-8), whereas it is approximately 55% for TCs (9-11).

The recommended curative treatment for localized TETs is complete surgical resection with or without postoperative radiation. However, there are little clinical data regarding second-line systemic therapy. For refractory or recurrent TETs, conventional chemotherapy has shown modest antitumor activity (12), while everolimus (13) for Ts and TCs, and sunitinib (14) and pembrolizumab (15) for TCs were accepted as second-line systemic therapies in the National Comprehensive Cancer Network (NCCN) guideline. However, due to the rarity of TETs, none of the targeted therapies or immunotherapies have been assessed in randomized phase 3 trials.

To enable discovery of novel targets and their potential clinical implications, we attempted to broadly and systematically investigate the comprehensive genomic alterations of TETs in a Chinese population to guide treatment decisions, especially for targeted therapy. Meanwhile, we also assessed the microsatellite stability (MSS) status, tumor mutation burden (TMB) value, and programmed death-ligand 1 (PD-L1) expression in TETs, which are predictive for immunotherapy.

We present the study in accordance with the MDAR reporting checklist (available at http://dx.doi.org/10.21037/gs-21-157).

**Methods**

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the ethical committee of Cancer Hospital of Shantou University Medical College (No. 2020061) and informed consent was taken from all the patients.

**Participants**

This study included 40 Chinese TET patients whose tumor and matched blood specimens were randomly collected between 2014 and 2019 from multiple centers. Histological subtypes for each case were evaluated by pathologists in each hospital according to WHO criteria. The diagnosis of the slides was reviewed by independent pathologists. All participants were clearly informed, understood the contents of the study, and agreed to the data of the study being published. Informed written consent was provided by each participant.

**Sample preparation**

Formalin-fixed paraffin-embedded (FFPE) tissue samples and matched blood specimens of TET participants were collected from accredited clinical hospitals. The histologic sections were retrieved. It was ensured that the percentage of tumor cells for each sample was more than 20%.

**Targeted next-generation sequencing (NGS) and genetic analysis**

Genomic DNA extraction (including FFPE tumor samples and matched white blood cells) and Cancer Sequencing YS panel (CSYS) was performed as previously described (16,17). Briefly, 50–250 ng DNA was extracted for subsequent genetic analysis. Genetic alteration detection was performed with CSYS panel which covers all the coding exons of 450+ cancer-related genes and selected introns of 39 genes that were frequently rearranged in solid tumors. Customized bioinformatics pipelines were applied for detection of single nucleotide variants (SNVs), short and long indels, copy number variations (CNVs), gene rearrangements, TMB, and microsatellite instability (MSI). All detected mutations were compared with our in-house database of genomic changes specific to clinical annotation.

**TMB calculation**

The TMB was estimated as previously described (17). Briefly, the total number of somatic mutations including coding SNVs and short indels were counted, and driver mutations and known germline alternations in the Single Nucleotide Polymorphism Database (dbSNP) were excluded. The TMB was calculated by dividing the total number of mutations counted by the size of the coding region.

**PD-L1 immunohistochemistry (IHC) staining assay**

The expression status of PD-L1 was estimated by IHC.
staining of FFPE tissue sections using anti-PD-L1 antibodies [clone 28-8; Cat#ab205921; Abcam (Cambridge, UK) or clone 22C3; Cat#M3653; Dako (Hovedstaden, Denmark)]. The specimen was considered to be PD-L1 positive if the staining cell percentage ≥1%.

Statistical analysis

Statistical analyses were performed using SPSS version 22.0 (SPSS Inc., Chicago, IL, USA). Fisher's exact test was used to analyze significant differences. P<0.05 was considered statistically significant.

Results

Demographic and clinicopathological characteristics of participants

To analyze the mutational features of TETs, a total of 40 patients were included in this study, including 21 Ts, 15 TCs, and 4 TNETs. Histological subtypes for each sample were evaluated by WHO criteria. The median age of participants at the time of sampling was 50.8 years (26–74 years), and 65% of participants were male. The demographic and clinicopathological characteristics of the 40 participants are shown in Table 1.

Landscape of genetic alterations

To investigate the distinctive genomic features of Chinese TETs, the 40 participants were subjected to a hybridization capture-based NGS panel (CSYS panel). Overall, 130 genetic alterations were identified in 84 genes. All participants had a signature of MSS except for 1 which was undefined. The average TMB was 2.285; approximately 1.75, 3.22, and 1.6 in Ts, TCs, and TNETs, respectively. The prevalence and distribution of genomic alterations are summarized in Figure 1.

The average number of mutations per participant was 3.25; approximately 1.43, 5.73, and 3.5 in Ts, TCs, and TNETs, respectively. Correlating with the lowest mutational prevalence, 4 out of 21 thymoma participants were identified with 0 mutations (case T03, T06, T08, and T19). Mutational analysis did not identify any recurrently mutated genes except for TAF1 and SETD2 which occurred in 2 cases (9.5%) in this subset. The scattered mutation distribution of Ts in a profiling map indicated the low recurrent frequency of the mutated genes. The

Table 1 Clinicopathological characteristics of 40 TET participants

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>n (%)</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Total number</td>
<td>40</td>
<td></td>
</tr>
<tr>
<td>Age (years), median [range]</td>
<td>50.8 [26–74]</td>
<td></td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>26 (65)</td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>14 (35)</td>
<td></td>
</tr>
<tr>
<td>Smoking status</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-smoker</td>
<td>16 (40)</td>
<td></td>
</tr>
<tr>
<td>Smoker and ex-smoker</td>
<td>10 (25)</td>
<td></td>
</tr>
<tr>
<td>Undefined</td>
<td>14 (35)</td>
<td></td>
</tr>
<tr>
<td>Stage</td>
<td></td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>4 (10)</td>
<td></td>
</tr>
<tr>
<td>II</td>
<td>0 (0)</td>
<td></td>
</tr>
<tr>
<td>III</td>
<td>6 (15)</td>
<td></td>
</tr>
<tr>
<td>IV</td>
<td>15 (37.5)</td>
<td></td>
</tr>
<tr>
<td>Undefined</td>
<td>16 (37.5)</td>
<td></td>
</tr>
<tr>
<td>Histological subgroup</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Thymoma (total 21)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Type A</td>
<td>1 (2.5)</td>
<td></td>
</tr>
<tr>
<td>Type AB</td>
<td>3 (7.5)</td>
<td></td>
</tr>
<tr>
<td>Type B1</td>
<td>1 (2.5)</td>
<td></td>
</tr>
<tr>
<td>Type B2</td>
<td>6 (15)</td>
<td></td>
</tr>
<tr>
<td>Type B3</td>
<td>4 (10)</td>
<td></td>
</tr>
<tr>
<td>Combined B</td>
<td>6 (15)</td>
<td></td>
</tr>
<tr>
<td>Thymic carcinoma (total 15)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Squamous cell carcinoma</td>
<td>10 (25)</td>
<td></td>
</tr>
<tr>
<td>Adenocarcinoma, NOS</td>
<td>1 (2.5)</td>
<td></td>
</tr>
<tr>
<td>Lymphoepithelioma-like carcinoma</td>
<td>1 (2.5)</td>
<td></td>
</tr>
<tr>
<td>Unknown</td>
<td>3 (7.5)</td>
<td></td>
</tr>
<tr>
<td>Thymic neuroendocrine tumor (total 4)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Carcinoid tumor</td>
<td>2 (5)</td>
<td></td>
</tr>
<tr>
<td>Small-cell carcinoma</td>
<td>1 (2.5)</td>
<td></td>
</tr>
<tr>
<td>Undefined</td>
<td>1 (2.5)</td>
<td></td>
</tr>
<tr>
<td>Autoimmune disease (in thymomas)</td>
<td>7 (33.3)</td>
<td></td>
</tr>
<tr>
<td>Myasthenia gravis</td>
<td>6 (28.6)</td>
<td></td>
</tr>
<tr>
<td>Systemic lupus erythematosus</td>
<td>1 (2.5)</td>
<td></td>
</tr>
</tbody>
</table>

TET, thymic epithelial tumor.
The most commonly mutated genes in TCs were CDKN2A (9, 60% in TC), CYLD (6, 40%), CDKN2B (5, 33.3%), and TP53 (4, 26.7%). Notably, the majority of mutations of CDKN2A were potential loss-of-function mutations, with the most significant gene deletions in 5 samples (case TC01, 03, 05, 11, and 14), missense mutations in 2 samples (case TC02 and 08), nonsense mutation in 1 sample (case TC13), and long deletion in exon 1 to intron 1 (case TC04). The genomic location of CDKN2A is at chr9q21.3, where CDKN2B and MTAP also locate. All 5 gene deletions of CDKN2A were accompanied by CDKN2B gene deletions, among which 3 cases harbored MTAP gene deletions (case TC01, 05, 11). TCs were characterized by gene deletions/amplifications, and 9 of the 15 samples had CNV of specified genes. For TNETs, we also observed scattered mutation distributions with no recurrently mutated genes identified except for MEN1. Case TN01 harbored 2 frameshift mutations in MEN1 (1 was somatic, the other was germline), and case TN03 harbored a somatic missense mutation in MEN1.

Given the prevalence and important role of MEN1 in neuroendocrine tumors, it was not surprising to observe the mutations in our subset.

**Predictive biomarkers with therapeutic perspective**

We explored the prevalence of actionable mutations of which targeted inhibitors have been encountered in clinical practice in TETs or other tumor types. Actionable mutations were based on databases including Oncology Knowledge Base (OncoKB, www.oncokb.org), Catalogue of Somatic Mutations in Cancer (COSMIC, cancer.sanger.ac.uk/cosmic), ClinicalTrials (cancer.sanger.ac.uk/cosmic/), and PubMed (www.ncbi.nlm.nih.gov/pubmed). Overall, 22 actionable mutations were identified in 8 genes. The tumor types with the highest proportion of actionable mutations were TCs (19, 86.4%), TNs (2, 9.1%), and Ts (1, 4.5%), sequentially. The most frequently recurrent actionable alterations included CDKN2A (n=9), CDKN2B (n=5), STK11 (n=2),
and DDR2 (n=2) (Figure 2). Altogether, 12 participants (30%), including 9 TCs, harbored at least 1 potentially actionable mutation.

IHC detection of PD-L1

Expression patterns of PD-L1 by IHC staining are depicted in Table 2. Inconsistent with a prior report (18), PD-L1 expression was predominant in males and smokers. The expression of PD-L1 was present in 50% (12/24) of specimens examined, including 54.6% of Ts, 54.6% of TCs, and 0% of TNETs.

Discussion

The incidence of thymic tumors is under-estimated, and despite rarity, TETs are aggressive malignancies. With the increasing use of computed tomography (CT) screening for lung cancer, more early stage TETs are likely to be discovered. Moreover, the different incidence of TETs between Chinese and Western populations suggests that there may be a genetic component (3,4). However, due to the rarity of TETs, there are few options for second-line systemic therapy. Therefore, precise therapeutic strategies with efficacy are needed.

Previous study showed that the most frequent mutated gene is GTF2I, and followed with HRAS, NRAS, and TP53.

Table 2 Overall expression patterns of PD-L1

<table>
<thead>
<tr>
<th>Clinical characteristic</th>
<th>PD-L1 expression (≥1+ by IHC), n (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>All (n=24)</td>
<td>12 (50)</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
</tr>
<tr>
<td>Male (n=15)</td>
<td>9 (60)</td>
</tr>
<tr>
<td>Female (n=9)</td>
<td>3 (33.3)</td>
</tr>
<tr>
<td>Smoker</td>
<td></td>
</tr>
<tr>
<td>Non-smokers (n=10)</td>
<td>4 (40)</td>
</tr>
<tr>
<td>Smoker and ex-smoker (n=7)</td>
<td>6 (85.7)</td>
</tr>
<tr>
<td>Histology</td>
<td></td>
</tr>
<tr>
<td>Thymomas (n=11)</td>
<td>6 (54.6)</td>
</tr>
<tr>
<td>Thymic carcinomas (n=11)</td>
<td>6 (54.6)</td>
</tr>
<tr>
<td>Thymic neuroendocrine tumors (n=2)</td>
<td>0 (0)</td>
</tr>
</tbody>
</table>

PD-L1, programmed death-ligand 1; IHC, immunohistochemistry.
in 117 TET patients, including 105 Ts, 10 TCs, and 2 microrodular thymomas, and most of patients were white race (19). Herein, we performed DNA targeted sequencing of cancer associated genes to analyze 40 Chinese TET patients. The results revealed that Ts, TCs, and TNETs are molecularly different malignancies. The mutational profiling of Ts and TNETs showed scattered mutation distributions with no recurrently mutated genes. In contrast, TCs did show highly recurrent mutations including CDKN2A, CYLD, CDKN2B, and TP53. The small number of TETs are the limitation of these studies, which still need to be further confirmed.

C-KIT overexpression is reported to associate with the worse prognosis of TET, and activating mutations of c-Kit is the biomarker associated with response to imatinib (20,21). Notably, we did not detect the c-KIT mutation in our TCs cohort for whom sunitinib is recommended. Previous studies reported the copy number aberrations of CDKN2A/B (22) and loss of p16INK4A (23) are associated with worse prognosis in TCs. The genes CDKN2A and CDKN2B lie adjacently at chr9p21.3 which is frequently mutated or deleted across a wide variety of tumors. The CDKN2A gene encodes p16INK4a and p14arf, while CDKN2B encodes p15Ink4b. The encoded proteins bind to CDK4/6 thereby inhibiting its kinase activity. Gene deletions or mutations of CDKN2A/B could lead to the activation of CDK4/6 (24). Therefore, CDK inhibitors might target the CDKN2A/B gene deletions or mutations. The Food and Drug Administration (FDA) has approved the CDK4/6 inhibitors, palbociclib, ribociclib, and abemaciclib, to treat hormone receptor (HR)-positive and human epidermal growth factor receptor receptor 2 (HER2)-negative advanced or metastatic breast cancer. Several studies indicated CDKN2A/B gene deletions or mutations patients could benefit from CDK4/6 inhibitors (25–28). Previous clinical trial (NCT01291017) showed that nearly half of advanced non-small cell lung cancer patients with wild-type RB and inactive CDKN2A, achieved stable disease for more than 4 months with the treatment of palbociclib.

Given pembrolizumab is active as second-line therapy in patients with TCs, we assessed the MSS status, TMB value, and PD-L1 expression in TETs. Overall, stable MSS and relatively low TMB value were observed in all 3 subtypes. Fortunately, 50% of Ts and TCs had PD-L1 expression which might be an indicative marker for immunotherapy in such patients.

Conclusions

In summary, we identified distinct genomic mutational profiles in Chinese TET patients with potentially actionable genetic alterations. The small number of patients is the limitation of this study. On account of the rarity and heterogeneity of TETs, it is critically important to initiate global or regional collaboration so as to accelerate the novel efficacy targeted- and immunotherapy in TETs.

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Footnote

Reporting Checklist: The authors have completed the MDAR reporting checklist. Available at http://dx.doi.org/10.21037/gs-21-157

Data Sharing Statement: Available at http://dx.doi.org/10.21037/gs-21-157

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/gs-21-157). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the ethical committee of Cancer Hospital of Shantou University Medical College (No. 2020061) and informed consent was taken from all the patients.

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