Introduction

Primary adrenal tumors comprise two main distinct types. Adrenal cortical tumors are derived from the cortex and mainly include adrenal cortical adenoma and cortical carcinoma. Adrenal tumors originating from the medulla contain pheochromocytoma (PCC) and neuroblastic tumors (NT). PCC and extra-adrenal paraganglioma (EAPGL) are both the catecholamine-secreting neuroendocrine tumors while the latter arise from the extra-adrenal paraganglia.

AQP2 as a diagnostic immunohistochemical marker for pheochromocytoma and/or paraganglioma

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Contributions: (I) Conception and design: D Niu, Y Bai; (II) Administrative support: D Niu, L Zhou; (III) Provision of study materials or patients: Y Bai, L Zhou, X Huang, C Zhao; (IV) Collection and assembly of data: D Niu, Q Yao; (V) Data analysis and interpretation: D Niu, Y Bai, Q Yao; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

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Background: Aquaporin2 (AQP2) is water channel protein that is widely distributed among mammalian tissues and plays a major role in water homeostasis. However, little is known about the expression and significance of AQP2 in human adrenal tumors. Thus, we performed an immunohistochemical investigation of AQP2 in normal and neoplastic adrenal tissues.

Methods: AQP2 protein expression was investigated in 190 adrenal tumor patients using immunohistochemistry. Correlation between protein expression and clinicopathological features was statistically analyzed.

Results: We demonstrated immunopositivity for AQP2 in all adrenal medulla-originating tumors, including 96 pheochromocytomas (PCC), 7 metastatic pheochromocytomas (MPCC), and 10 neuroblastic tumors (NT) and 13 extra-adrenal paragangliomas (EAPGL). Whereas, APQ2 was absent from the 52 adrenal cortical adenomas and 2 adrenal cortical carcinomas examined. The 10 metastatic carcinomas examined in adrenal tissue were also negative for AQP2. In 82 (85%) of the 96 samples from patients with PCC, we studied the relationship between clinicopathologic factors and AQP2 expression and our findings suggested that the tumors that exhibited diffuse expression pattern of AQP2 were larger in diameter than those exhibiting focal (P=0.007) or mediate expression pattern (P=0.001).

Conclusions: AQP2 protein is significantly expressed in normal adrenal medullary tissues and medullary tumors (including PCC, MPCC and NT) as well as EAPGL. AQP2 expression may indicate the origin of normal adrenal tissues, and its expression in cancer tissue may reflect the maintenance of water metabolism via AQP2 during tumorigenesis. AQP2 may serve as a valuable marker for the differential diagnosis of adrenal tumors.

Keywords: Aquaporin2 (AQP2); immunohistochemistry; pheochromocytoma (PCC); paraganglioma

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About ~40% of PCCs and EAPGLs are identified as inherited forms due to the germline mutation in susceptibility genes such as \( VHL \), \( SDHB \), \( SDHD \) and \( SDHC \) genes. The remaining 60% are thought to be sporadic cases. At least one-third of the sporadic tumors contain a somatic mutation in predisposing gene including \( VHL \), \( RET \), \( NF1 \) and \( HRAS \) (1,2).

The aquaporins (AQPs) are a family of small membrane proteins (25~34 kDa) of 13 subtypes (AQP0–AQP12) that serve as hydrophobic and integral membrane channel proteins that facilitate rapid passive movement of water (3). AQPs not only play important roles in fluid transport in many epithelial and endothelial tissues, but they also participate in the regulation of important cellular functions, including cell proliferation and migration, apoptosis, phagocytosis and neuronal signal transduction (4). In neoplastic tissues, AQP subtypes were involved in invasion, migration, and proliferation of tumors in several organs, including skin, urinary bladder, stomach, colon, kidney (5-8) and so on. Among these subtypes, AQP2 is a classical water channel protein that regulates water homeostasis in normal tissue (9), and its expression has been demonstrated in the kidney cortex/medulla, gastric pit, small intestine, pancreatic islet, and fallopian tubes, among other tissues (10). AQP2 is responsible for regulating the volume and osmolarity of urine in the kidney (10) and involves pain transmission in the trigeminal nerve (4). Recently, the expression of AQP2 was reported in endometrial carcinoma and brain glioma, where it mediates estrogen-induced cell migration and invasion (11,12). Very limited information is available concerning the role of AQP2 in human adrenal tumors.

In the current study, we performed immunohistochemical analysis of AQP2 expression in 190 adrenal tumor samples. Our findings implicated AQP2 as a valuable indicator for differential diagnosis of adrenal tumors.

### Methods

#### Cases selection

We selected a series of surgical specimens of neoplastic adrenal tissues (190 cases), which were consecutively collected between 1993 and 2012 from Peking University Cancer Hospital & Institute. The cases, summarized in Table 1, included adrenal cortical adenoma (n=52), adrenal cortical carcinoma (n=2), PCC (n=96), metastatic pheochromocytoma (n=7), EAPGL (n=13), NT (n=10), and metastatic carcinoma (n=10). We also investigated normal tissue adjacent to the tumors. Hematoxylin-eosin (HE) staining was carried out on all of the slides, and the corresponding diagnosis was made on the basis of the World Health Organization classification criteria (13). All MPCC cases had metastasized to lymphatic nodes and/or several organs including liver, lung, pancreas, and colon. All patients with metastatic carcinoma had a history of primary cancer, including 2 hepatocellular carcinomas, 2 renal cell carcinomas, 3 pulmonary carcinomas, 1 breast carcinoma, 1

<table>
<thead>
<tr>
<th>Tumor types</th>
<th>AQP2 expression</th>
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<tbody>
<tr>
<td></td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>1+</td>
</tr>
<tr>
<td></td>
<td>2+</td>
</tr>
<tr>
<td></td>
<td>3+</td>
</tr>
<tr>
<td>Adrenal cortical tumor</td>
<td></td>
</tr>
<tr>
<td>Adrenal cortical adenoma (n=52)</td>
<td>52 (100%)</td>
</tr>
<tr>
<td>Adrenal cortical carcinoma (n=2)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Adrenal medullary tumor</td>
<td></td>
</tr>
<tr>
<td>Pheochromocytoma (n=96)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Metastatic pheochromocytoma (n=7)</td>
<td>10 (10%)</td>
</tr>
<tr>
<td>Neuroblastic tumor (n=10)</td>
<td>66 (60%)</td>
</tr>
<tr>
<td>Extra-adrenal paraganglioma (n=13)</td>
<td>65 (52%)</td>
</tr>
<tr>
<td>Other tumors</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Metastatic carcinoma (n=10)</td>
<td>10 (100%)</td>
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</tbody>
</table>

0, negative; 1+, focal (1% to 9%); 2+, intermediate (10% to 50%); 3+, diffuse (more than 50%).
rectal carcinoma and 1 gastric carcinoma. Clinicopathologic factors including age, sex, and tumor size were examined in 82 (85%) of the 96 patients with PCC. The study protocol was approved by the institutional ethics committee of the Peking University Cancer Hospital & Institute (No. 2014KT196).

**Immunohistochemical assay**

Immunohistochemical analysis was performed on 3-μm sections of formalin-fixed and paraffin-embedded tissues mounted on silanized slides. The deparaffinized sections were placed in plastic Coplin jars filled with 10 mmol/L citrate buffer (pH 6.0) and incubated for 10 minutes at 120 °C in an autoclave for heat antigen retrieval. After the autoclave treatment, endogenous peroxidase was quenched with 3% (vol/vol) H₂O₂ for 10 minutes. The sections were incubated with an anti-AQP2 antibody (1:1,000 dilution, Sigma, USA) at room temperature for 1 hour. We used normal tissue from the collecting duct of the kidney as a positive control and omitted the primary antibody for the negative control. We quantified the immunoreactivity of AQP2 using a four-tier scale from 0 to 3+: 0= no staining, 1+ = focal (1% to 9% positive cells), 2+ = intermediate (10% to 50% positive cells), 3+ = diffuse (50% or more positive cells). Immunohistochemical staining was analyzed by two observers (DN and YB) blinded to all clinicopathological information. Discrepancies in immunohistochemical analysis were reconciled following the discussion of 2 observers.

**Statistics**

The clinicopathologic characteristics were statistically analyzed by ANOVA (age, tumor size, and tumor weight versus AQP2 positive pattern) and multiple comparison tests (sex versus AQP2 positive pattern). We set statistical significance at P<0.05 and performed data analyses with SPSS V.25.0 (SPSS, Chicago, Illinois, USA).

**Results**

**AQP2 in normal tissues**

We chose normal kidney tissues as a positive control. As expected, AQP2 protein was localized specifically in the membrane of the collecting duct cells of the kidney (Figure 1). Then we studied the immunostaining of AQP2 in adrenal tissue and observed immunopositivity of AQP2 in cells of the adrenal medulla rather than in adrenal cortical cells (Figure 2).

![Figure 1](image1.png) **Figure 1** Immunohistochemical staining of AQP2 in normal kidney tissue for positive control. The cytoplasmic membrane of collecting ducts presents immunopositive for AQP2. (Magnification: ×600).

![Figure 2](image2.png) **Figure 2** Immunohistochemical staining for AQP2 in normal adrenal tissues/cells. (A) Normal adrenal medullar cells and cortical cells is observed (H&E, magnification: ×200); (B) diffuse immunopositive for AQP2 is showed in adrenal medullar cells, but not in cortical cells (magnification: ×200).
AQP2 expression in adrenal PCC and EAPGLs

The immunohistochemical results of AQP2 for adrenal PCC and EAPGLs are summarized in Table 1. We found that AQP2 protein was expressed in all adrenal medullary tumors and EAPGLs. Among 96 PCCs, most tumors exhibited a diffuse pattern of AQP2 expression, identified in 85% (81/96) of cases, while focal expression and intermediate expression were observed in only 5% (5/96) and 10% (10/96) of cases, respectively. Immunohistochemical intensity of AQP2 in PCC tumors with Zellballen architecture (Figure 3A,B) was notably strong compared to tumors with solid architecture (Figure 3C,D). In MPCC tissues, the distribution of AQP2-positive cells varied: 57% (4/7) in diffuse, 29% (2/7) in intermediate and 14% (1/7) in focal tumor tissues (Figure 4A,B,C,D,E,F). The focal expression pattern of AQP2 was identified in 4 (40%) of 10 Schwannian stroma cells of NTs. All 13 EAPGLs exhibited a diffuse distribution of AQP2.

AQP2 expression pattern in adrenal cortical tumors and other adrenal tumors

We found that AQP2 was not expressed in the adrenal
cortical tumors, which included 52 cortical adenomas and 2 cortical carcinomas (Figure 5A,B,C,D). All 10 metastatic carcinomas were also negative for AQP2 protein.

The association between clinicopathologic factors and AQP2 expression in adrenal PCCs

The patients with PCC comprised 44 women and 38 men, ranging from 29 to 83 years in age (median, 50). The tumor diameter ranged from 1.1 to 45 cm (median, 5). The tumor weight ranged from 5 to 740 g (median, 45). The tumors showing the diffuse expression pattern of AQP2 were larger in diameter than those with a focal (P=0.007) or mediate expression pattern (P=0.001). Besides, there were no significant effects of other clinicopathologic factors (age, sex, and tumor weight) on the expression of AQP2. All data was showed in Table 2.

Discussion

AQP2 is a typical water channel protein that plays an essential role in fluid homeostasis in normal tissues. AQP2

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Table 2 Clinicopathologic factors and their relation with AQP2 expression

<table>
<thead>
<tr>
<th>Clinicopathologic factors</th>
<th>1+</th>
<th>2+</th>
<th>3+</th>
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<tbody>
<tr>
<td>Age (years)</td>
<td>50.4±11.3</td>
<td>51.8±11.3</td>
<td>52.7±14.0</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Male (n=38) (%)</td>
<td>4.8</td>
<td>7.1</td>
<td>88.1</td>
</tr>
<tr>
<td>Female (n=44) (%)</td>
<td>6.3</td>
<td>10.4</td>
<td>83.3</td>
</tr>
<tr>
<td>Tumor size (cm)</td>
<td>6.1±5.6</td>
<td>5.8±2.6</td>
<td>17.4±16.0**</td>
</tr>
<tr>
<td>Tumor weight (g)</td>
<td>48.9±38.7</td>
<td>68.0±51.1</td>
<td>101.0±72.6</td>
</tr>
</tbody>
</table>

0, negative; 1+, focal (1% to 9%); 2+, intermediate (10% to 50%); 3+, diffuse (more than 50%). Statistical significance was calculated using the ANOVA test (age, size, and weight) and multiple comparison (sex). **, P<0.01.

Figure 5 Immunohistochemical staining for AQP2 in adrenal cortical tumor tissues. Adrenal cortical adenoma presents microvesicle or clear cytoplasm mimicking the zona fasciculata (A). Adrenal cortical carcinoma shows severe nuclear atypia, and interstitial myxoid change (C) (H&E, magnification: ×200). The tumor tissues/cells are negative for AQP2 (B,D, magnification: ×400).

was originally identified in the renal collecting duct cells, where it functions as the vasopressin-sensitive water channel (14-16). Its expression also has been reported in pancreatic islets, fallopian tubes, peripheral/central nerves, and gastrointestinal tissues (10).

To our knowledge, the expression of AQP2 has not been reported in normal adrenal gland tissues. In this study, we used immunohistochemical analysis to show that AQP2 protein was expressed in normal adrenal medullary cells but not in adrenal cortical cells. It has been confirmed that AQP2 is linked to vasopressin-stimulated water reabsorption in kidney (17). From this point of view, it is
conceivable that AQP2 in adrenal medullar cells may play an important role in water homeostasis and other biological functions. However, its expression also raises two questions: (I) is AQP2 the only AQP subtype with a functional role in adrenal medullar cells; and (II) which subtypes of AQP are responsible for water transport in adrenal cortical cells? We previously demonstrated that AQP3 might play a role in fluid homeostasis in medullary thyroid carcinoma (18) but not in adrenal gland tumors (19). However, additional studies are needed to further elucidate these questions. Moreover, in embryology, the medulla and cortex of the adrenal gland have distinct embryonic origins. Medullary cells originate from the neuroectoderm, whereas cortical cells originate from the mesoderm. Thus, our findings also suggested that AQP2 expression may reflect the origin of normal adrenal gland tissues.

A lack of function and/or reduced expression of AQP2 has been observed in several diseases, such as renal failure (20), acquired nephrogenic diabetes insipidus, and post-obstructive polyuria (21). AQP2 can also induce the cellular migration and invasion of endometrial carcinoma (11). We demonstrated that AQP2 was specially expressed in all adrenal medulla-originating tumors but not in cortical-originating tumors. These findings suggested that AQP2 may be associated with the biological metabolism of water in PCCs. However, we failed to find any difference on AQP2 staining between PCC and MPCC, which implied that AQP2 may not be served as a valuable predictive parameter on metastasis disease. According to our findings, the immunohistochemical staining of AQP2 can be used as helpful diagnostic markers to distinguish PCCs and paraganglioma from other adrenal tumors such as adrenal cortical tumors and metatstatic tumors for routine pathologic diagnosis, similar to the neural markers chromograninA (13), among others. AQP2 expression in normal adrenal medullar tissues and in adrenal medulla-originating tumors also implies that the water metabolism function of AQP2 is well maintained during tumorgenesis.

We analyzed the correlation between AQP2 expression and clinicopathologic factors in adrenal PCCs. The results showed that tumors with the diffuse expression pattern of AQP2 were larger in diameter than those displaying focal or mediate expression patterns. AQP subtypes play important roles in cell growth and proliferation in several types of cancer, such as squamous cell carcinomas (22,23), breast carcinomas (24), ovarian carcinomas (25), and brain tumors (26). Thus, it is reasonable to suggest that AQP2 may be associated with the growth and/or proliferation of PCCs, and the present findings confirmed our speculation that AQP2 participates in the biological metabolism of water in the tumors. Because of the limited cases of MPCC, we failed to compare the AQP2 expression and clinicopathologic factors between PCC and MPCC. In addition, we only analyzed AQP2 immunohistochemical expression in adrenal tumors, but the AQP family comprises 13 subtypes with differential expression patterns in normal tissues. Different combinations of these various types of AQPs may be more valuable for diagnosis of adrenal tumors, warranting further studies.

In conclusion, the AQP2 protein is expressed in normal adrenal medullary tissues and in medullar tumors (PCC, MPCC and NT) as well as in EAPGL, but it is not found in normal adrenal cortical tissue or in cortical tumors. Thus, AQP2 may have differential diagnostic value in identifying adrenal tumors. In addition, PCCs with a diffuse pattern of AQP2 expression were larger in diameter than those displaying focal or mediate expression. Our findings also suggest that AQP2 expression may reflect the origin of normal adrenal gland tissues. AQP2 not only participates in the biological metabolism of water in PCC but also likely maintains this function during tumorgenesis in adrenal medullar tumors.

Acknowledgments

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/gs.2020.01.19). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The research program was approved by the ethics committee of Peking University Cancer Hospital & Institute (No. 2014KT196).
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